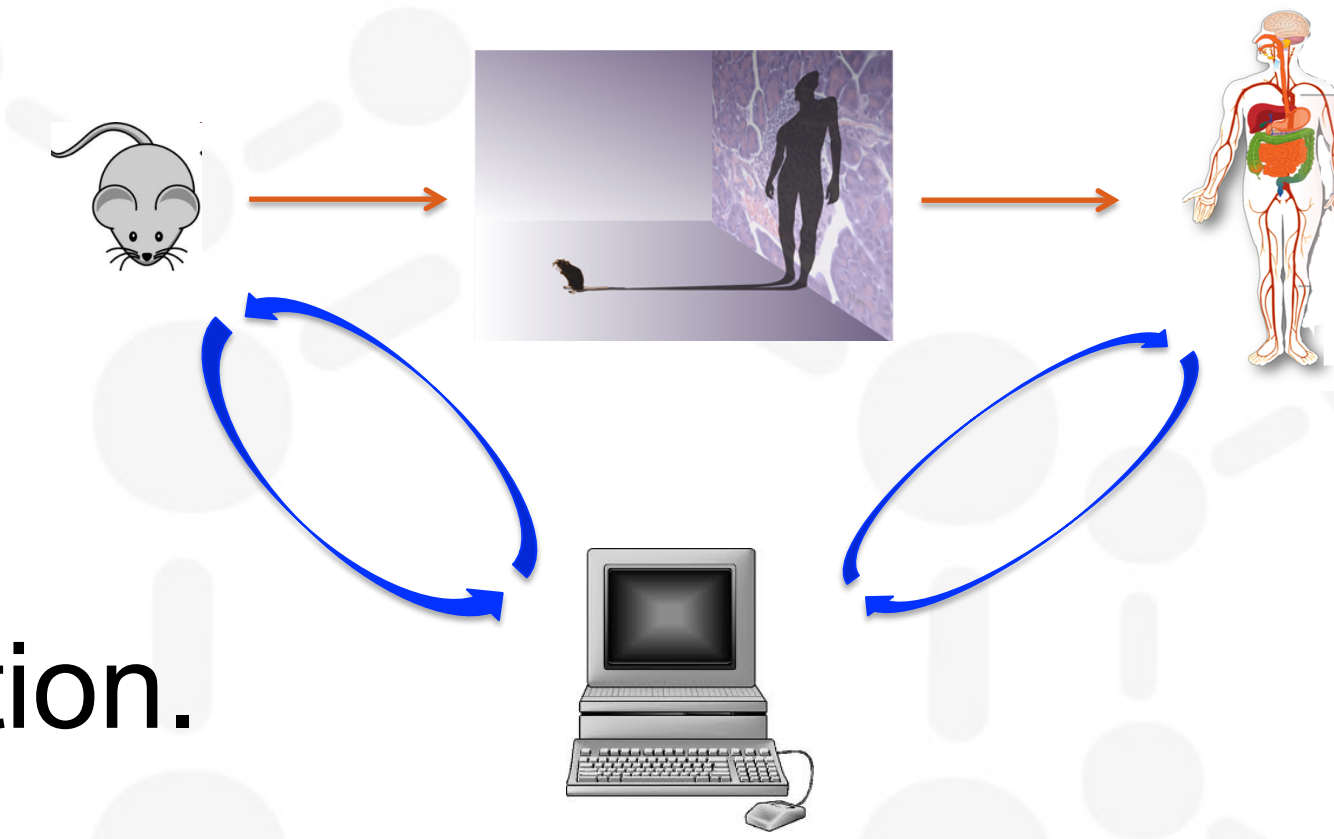


# Poster 17: LiSyM Midterm Evaluation 2018 Drasdo Group (Dortmund)

## Facts & Figures:

Area of Expertise: Modelling  
Methodes: Computational Multi-Cellular Agent-Based Modeling, Flow Simulation.  
Scientific Involvement: Pillar II, III  
LiSyM Resources: ~ 600 000 € / 3yrs, ~1 Mio € / 5yrs  
LiSyM Scientists: PhD Student: Noémie Boissier; PostDocs: Jieling Zhao, Ayham Zaza, Paul van Liedekerke; Research Engineer: Tim Johann  
Collaboration (main): Groups Hengstler (Dortmund), Höhme (Leipzig), Dooley (Mannheim), Klingmüller, Kauczor (Heidelb), Timmer (Freiburg); Junior group Ghallab(Do)



**Objectives: general**

- Development of mathematical models to guide experimental designs & clinical decisions
- Long-term prospective: virtual twin of a patient

**Objectives: specific**

- Fibrosis: disease progression
- Fibrosis: functional consequences
- ACLF: acute damage in advanced fibrosis = acute on chronic

**Acute damage: reference model for ACLF & chronic damage**

Liver lobule by confocal microscopy using image processing pipeline (TiQuant -> Poster 3)

**Model setup: APAP-regeneration damage & HGF – control of regeneration**

- Multi-scale model, executing APAP & HGF-pathway in each hepatocyte
- Each cell represented within "center-based"-model that mimics forces between cells as forces between cell centers
- Cells move due to an equation of motion (one per cell) as consequence of active and passive forces
- Cell division controlled by HGF-controlled cell cycle progression
- Cell death below ATP threshold concentration triggered by APAP
- Blood flow and molecular transport modeled as for ammonia detoxification (see below)
- Coupling to extra-hepatic body model for HGF clearance and APAP pharmacokinetics by population PK or PBPK model

**Results:**

**APAP: simulation scenario**

**Regeneration: simulation scenario**

Model permits extrapolation from in-vitro -> in-vivo for APAP  
Model explains data on regeneration after APAP damage quantitatively

**Models: functional consequence: ammonia detoxification**

**Poiseuille flow**

$$Q^{(ij)} = v^{(ij)} \pi \cdot (R^{(ij)})^2 = -\frac{\pi \cdot (R^{(ij)})^4}{8\eta(R^{(ij)})L^{(ij)}} (p^{(i)} - p^{(j)})$$

**Transport: mass balance**

$$V_{\text{Hep}} \partial_t c_{\text{H}} + \nabla \cdot (v_{\text{Hep}} c_{\text{H}} V_{\text{Hep}}) = -K_{\text{Hep}} (c_{\text{H}} - c_{\text{B}})$$

**Change of mass in sinusoid**  $\rightarrow$  **Transport to hepatocyte**

**Change of mass in hepatocyte**  $\rightarrow$  **Transport from sinusoid**  $\rightarrow$  **Metabolism**

Model-guided identification of an unrecognized ammonia sink (reversible GDH-reaction)  
Reaction can be triggered in blood and saves mice from hyperammonemia

All simulations base on implementation in own software TiSim & TiQuant [contact Tim Johann](#)

**Fibrosis after CCl4: impact of architectural distortion on ammonia detoxification**

Sirius red staining of fibrotic tissue: bright field micrographs, whole slide scans

**Fibrotic tissue: confocal micrographs**

**Models: functional consequence: ammonia detoxification**

- Reduced permeability of sinusoids in fibrotic walls alone leads to reduced capacity of liver to detoxify blood from ammonia.
- For chosen geometries, no impact on flow.

**Fibrosis: Chronic CCl4 model: disease progression: CYP & ECM**

**Modeled CYP pattern. Sensitivity ~ 93%, but false positive**

**CYP2E1 pattern: bridging after repetitive CCl4 exposure**

**PSR: formation of fibrotic walls along CYP-positive pattern**

**CYP-pattern: Activator-inhibitor system reproduces bridging in whole-slide scan:**

$$\frac{da}{dt} = s_{a,cv} - D_a \Delta a - d_a a - kab$$

$$\frac{db}{dt} = s_{b,pv} - D_b \Delta b - d_b b - gab$$

**Towards a model of fibrotic wall formation**

- Hypothesis: **After multiple doses**, removal not happening, leading to accumulation of ECM at lesion localizations that regenerate (close) latest.
- ECM modeled as network of springs with bending rigidity

**After single APAP-dose:**

- Hepatic stellate cells in CYP-pos region at loc. of max. damage
- Macrophages peak close to CV at 4d, disappearing after closure
- Suggests HCSs produce ECM, later removed by macrophages

**Architectural distortion after Western diet and repetitive CCl4-doses: call for new model**

**Center-based model cannot capture correctly architecture distortion & biomechanics after WD or CCl4-induced fibrosis -> Deformable Cell Model (DCM), triangulating cell shape. -> DCM calibrated with optical stretcher experiment**

**Results:**

Regeneration after APAP-induced damage occurs at lower forces in DCM than for center based model, as cells in DCM naturally deform.

**Publications (only those appeared):** Ghallab, Celliere, ..., Drasdo\*, Gebhardt\*, Hengstler\* 2016. Journal of Hepatology 64(4): 860 (\*shared senior authors); Hoehme\*, Bertaux\*, ... Hengstler, Drasdo\*, 2018 (\*shared first authors). Bull. Math. Biol. Volume 80, Issue 5; Leist et. al. Arch Toxicol. 2017 Nov;91(11); Peters et. al. J Anat. 2017 Mar;230(3):471-483; Hoehme, ..., Drasdo, Hengstler, Methods Mol Biol. 2017;1506:319-362